

'6 β -PROPANOYLOXY-3 α -TIGLOYLOXYTROPANE, A NEW ALKALOID FROM *DATURA INNOXIA*

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Abstract—A new alkaloid, shown by spectroscopic and degradative means to be 6 β -propanoyloxy-3 α -tigloyloxytropane has been isolated from *Datura innoxia* root. The synthesis of the natural and (\pm) base is described.

Datura innoxia Miller (Solanaceae) is a herbaceous white flowered annual which is classified along with the closely similar *D. meteloides* DC ex Dunal in Section Dutra of the genus.¹ It has been extensively investigated in the past and has been the source of several new tigloyl esters.^{2,3} During routine analysis of the root for 3 α ,6 β -ditigloyloxytropane and 3 α ,6 β -ditigloyloxytropan-7 β -ol using the modified³ Evans and Partridge system,⁴ a small shoulder on the elution curve was noticed on the down-side of the 3 α ,6 β -ditigloyloxytropane peak. Although present in only a small quantity (0.003%) the new base, $[\alpha]_D^{25} = 0^\circ$, picrate m.p. 163° was isolated by bulking together material from several columns. The base C₁₆H₂₅NO₄ (MS) had M = 295 (6%) and a fragmentation pattern characteristic of the tropane nucleus.⁵ Since the base peak was at $m/e = 94$ it was assumed that the alkaloid was a dihydroxytropane derivative, and the peak at $M^+ - 99$ was tentatively assigned to the loss of a tigloyl residue. The IR showed two ester carbonyl bands at 1740 and 1708 cm⁻¹ (unsat.), and >C=C< at 1630 cm⁻¹. Examination of the NMR spectrum (in CDCl₃ using TMS as internal standard) confirmed the presence of a di-substituted tropane nucleus with one tigloyl group,^{6,7} and showed the following signals: $\tau = 4.49$ (*dd* due to the tropane C(6) proton coupling with the C(7) α and β protons); 4.89 (*t* tropane 3 β proton) thus indicating the α orientation for the C(3) acyl group;⁸ 6.76 [*bm* tropane C(1) and (5)]; 7.43 (*s* *N*-Me); 3.10 (*q* tigloyl 3 β proton coupling with the β methyl); 8.13 (*s* tigloyl α methyl); 8.17 (*d* tigloyl β methyl partly overshadowed by the signal at $\tau = 8.13$). In addition signals at $\tau = 7.67$ (*q* MeC $\underline{\text{H}}$ CH₂) and 8.86 (*t* C $\underline{\text{H}}$ CH₃CH₂) suggested that the base was probably a propanoyl ester. Hydrolysis of the base gave two acids, propanoic and tiglic, identified by GLC and in the case of tiglic acid m.p. and m.m.p. The alkaline was esterified

¹ BLAKESLEE, A. F. (1959) *The Genus Datura* (AVERY, A. G., SATINA, S. and RIETSEMA, J., Eds.), p. 16, Ronald Press, New York.

² EVANS, W. C. and WELLENDOFF, M. (1958) *J. Chem. Soc.* 1991.

³ EVANS, W. C. and WELLENDOFF, M. (1959) *J. Chem. Soc.* 1406.

⁴ EVANS, W. C. and PARTRIDGE, M. W. (1949) *J. Pharm. Pharmacol.* 1, 593.

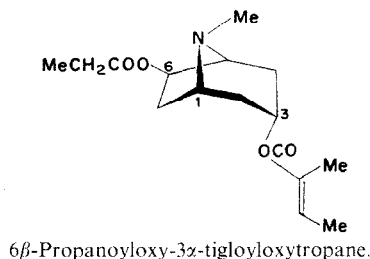
⁵ BLOSSEY, E. C., BUDZIKIEWICZ, H., OHASHI, M., FODOR, G. and DJERASSI, C. (1964) *Tetrahedron* 20, 585.

⁶ EVANS, W. C. and MAJOR, V. A. (1966) *J. Chem. Soc.* 1621.

⁷ BISHOP, R. J., FODOR, G., KATRITSKY, A. R., SOTI, F., SUTTON, L. E. and SWINBOURNE, F. J. (1966) *J. Chem. Soc.* 74.

⁸ PARELLO, J., LONGEVIALE, P., VETTER, W. and McCLOSKEY, J. A. (1964) *Bull. Soc. Chim. Fr.* 2787.

with tigloyl chloride and gave (–)-3 α ,6 β -ditigloyloxytropane thereby indicating that it was (+)-tropan-3 α ,6 β -diol which is known to have the 3R 6R configuration⁹ (structure).



The relative positions of the tigloyl and propanoyl moieties were determined in two ways. Partial hydrolysis^{10,11} of (–)-3 α ,6 β -ditigloyloxytropane gave (–)-6 β -hydroxy-3 α -tigloyloxytropane in 34% yield. Esterification of the latter base with propanoyl chloride gave a product indistinguishable from the natural base (slightly higher picrate m.p.). Secondly, (±)-6 β -hydroxytropane-3-one was esterified with propanoyl chloride and reduced to the 3 α -ol using Raney nickel. The 3 α -tigloyl ester had NMR, MS and IR spectra and R_f values on TLC and partition columns very similar to the natural alkaloid.

The appearance of a propanoyl ester in the Solanaceae tropane series seems to be unique, although 3 α -propanoyloxytropane has been isolated from *Bruguiera sexangula* and *B. exaristata* (Rhizophoraceae).¹² Similarly, the hetero di-esters of dihydroxytropane are extremely rare, 6 β -acetoxy-3 α -tigloyloxytropane and 6 β -(2-methylbutanoyloxy)-3 α -tigloyloxytropane occurring in *D. sanguinea* R. and P.,⁶ and *D. suaveolens* H. and B. ex Willd., respectively.¹³ Both these plants are in Section Brugmansia (perennial tree-daturas), a group quite different from the more common herbaceous daturas with perhaps the exception of *D. ceratocaula* Ort. The dextrorotatory enantiomer of dihydroxytropane appears to be a characteristic of the genus as a whole but (–)-tropan-3 α ,6 β -diol (valerine) occurs only in *Duboisia*¹⁴ as the alkamine of the alkaloid valeriodine.

EXPERIMENTAL

Optical rotations were measured using a Bendix NPL automatic polarimeter type 143. *Datura innoxia* plants were grown on open land in the University of Nottingham and were of the same strain (English sample) examined previously.³

Isolation of alkaloids. Dried, finely powered *D. innoxia* root (50 g) was mixed with 12.5 g Ca(OH)₂, moistened with H₂O (ca 25 ml) and exhaustively extracted with Et₂O. After removal of the solvent, the residue in CHCl₃ (ca 2 ml) was transferred to a kieselguhr column (10 g) supporting 0.5 M phosphate buffer pH 6.8 (5 ml). Elution with petrol. (100 ml) gave a mixture of bases which was further resolved on a kieselguhr column (10 g) containing 5 ml 0.5 M phosphate buffer pH 5.6. The elution curve (titre, ml 0.005 N H₂SO₄ using bromocresol green indicator vs fraction (5 ml) number) showed three peaks designated A, B and C: in petrol. b.p. 40–60, A (sharp) 24 mg; B (shoulder) 1.8 mg; in Et₂O, C (sharp) 10.1 mg. TLC Alumina (Merck), Et₂O, detected with I₂/CCl₄ suggested that A and C were 3 α ,6 β -ditigloyloxytropane and 3 α ,6 β -ditigloyloxytropane-7 β -ol respectively. These bases were characterized as the picrates² (m.p., m.m.p. and IR). The separation was repeated several times in order to isolate sufficient B. Base B, colourless gum, $[\alpha]_D^{25} = 0^\circ$ (undetectable). Picrate recryst. from EtOH/H₂O m.p. 163° (needles); IR $\nu_{\text{max}}^{\text{KBr}}$ 1740, 1708, 1630, 1560, 1485, 1430, 1365, 1318, 1245, 1150, 1065, 907, 780, 735, 705

⁹ FODOR, G. and SOTI, F. (1965) *J. Chem. Soc.* 6830.

¹⁰ FODOR, G., VINCZE, I., TOTI, J., JANZSO, G. and LANG, K. U.S.P. 2,905,687; B.P. 824, 623.

¹¹ EVANS, W. C. and GRIFFIN, W. J. (1963) *J. Chem. Soc.* 4348.

¹² LODER, J. W. and RUSSELL, G. B. (1969) *Aust. J. Chem.* **22**, 1271.

¹³ EVANS, W. C. and LAMPARD, J. F. (1972) *Phytochemistry* **11**, 3292.

¹⁴ BARGER, G., MITCHELL, W. and MARTIN, W. F. (1937) *J. Chem. Soc.* 1820.

cm^{-1} ; MS m/e with $I\%$ in parentheses, 42 (23), 55 (26), 57 (16), 81 (13), 82 (10), 83 (20), 94 (100), 95 (95), 96 (14), 122 (39), 138 (19), 195 (8), 196 (11), $M^+ = 295$ (6). Accurate mass measurement, $M^+ = 295.1787$, $\text{C}_{16}\text{H}_{25}\text{NO}_4$ requires 295.1784. NMR (base regenerated from the picrate in CDCl_3 using TMS as internal standard), $\tau = 3.10$ (1H q J 7 Hz), 4.49 (1H dd J 3 and 5 Hz), 4.89 (1H t J 4 Hz), 6.76 (bm), 7.43 (3H s), 7.67 (2H q J 8 Hz), 8.13 (3H s), 8.17 (3H d J 4 Hz), 8.86 (3H t J 7 Hz).

Hydrolysis of base. The base (4.5 mg) was heated at 100° in a sealed tube with 5% $\text{Ba}(\text{OH})_2$ (7 ml) for 2 hr. Cooled hydrolysate was acidified (80% H_2SO_4) and extracted with Et_2O (4×10 ml). The remaining aqu. phase was neutralized (solid BaCO_3), centrifuged and the supernatant liquid and washings were evaporated to dryness under reduced pressure. The dried residue was then heated under reflux with tigloyl chloride (0.2 ml) for 3 hr,¹⁴ cooled, washed several times with Et_2O , basified, and extracted with CHCl_3 . Evaluation of the product by TLC (Alumina, Et_2O) showed only one spot with an R_f identical to that of 3 α ,6 β -ditigloyloxytropone. The picrate (yield 2 mg) had m.p. and m.m.p. and IR identical to authentic (–)-3 α ,6 β -ditigloyloxytropone picrate, and the base recovered from the picrate had $[\alpha]_D^{25} = -18.46^\circ$ c.f. -18.82° determined on the authentic base under the same conditions. The Et_2O extract of the acidified hydrolysate was examined by GLC on a 3 ft celite column containing 10% silicone oil and 2% orthophosphoric acid; N flow rate 30 ml/min, F.I. detector with temp. programme 30–160° (32°/min).⁹ After GLC, the Et_2O was allowed to evaporate and this deposited a few crystals of tiglic acid m.p. 62° .

Partial synthesis of 6 β -propanoyloxy-3 α -tigloyloxytropone. (–)-3 α ,6 β -Ditigloyloxytropone (58 mg) in Me_2CO (1.5 ml) and 0.1 N NaOH 2.7 ml maintained at 22° for 4 hr, and then neutralized with HCl (10%). The Me_2CO was removed *in vacuo* and the remaining basified (NH_4OH) solution extracted with CHCl_3 . Evaporation of the CHCl_3 gave an oily straw-coloured residue which was resolved on a partition column at pH 6.8. Elution with petrol. (75 ml) gave unchanged starting material, Et_2O (75 ml) gave 6 β -tigloyloxytropone-3 α -ol¹¹ and CHCl_3 gave (–)-6 β -hydroxy-3 α -tigloyloxytropone (14.8 mg), which was dried and heated under reflux (80 – 85°) with propanoyl chloride (0.2 ml), for 2 hr. The product was purified by partition column chromatography at pH 6.8 (yield 10 mg) $[\alpha]_D^{25} = 0^\circ$, NMR identical with natural base, picrate m.p. 168° .

(\pm)-6 β -Propanoyloxy-3 α -tigloyloxytropone. Dry 6 β -hydroxytropone-3-one (25 mg) was esterified with propanoyl chloride (0.5 ml) and the recovered base in EtOH reduced (at. press.) with H_2 in the presence of Raney Ni. The product (TLC pure 6 β -propanoyloxytropone-3 α -ol R_f 0.59 Alumina $\text{Et}_2\text{O}:\text{EtOH}$ 4:1) was not isolated but esterified with tigloyl chloride and purified by the method outlined above to give (\pm)-6 β -propanoyloxy-3 α -tigloyloxytropone, NMR identical to natural base, picrate m.p. 169° .

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